# WHEAT GLUTEN, COELIAC DISEASE AND THE MICROBIOME: EXPLORING THE CONNECTIONS

CHARLES PAUL "MAX" MOEHS

Seattle, USA (e-mail: cpmmoe@gmail.com)

## **SUMMARY**

Coeliac Disease (CD) is the most common food-induced immune mediated enteropathy in humans with an estimated prevalence of approximately 1% in worldwide human populations. A related condition known as non-coeliac gluten sensitivity (NCGS) appears to be increasing in prevalence, and may be even more common in the general population. These diseases are induced by the consumption of gluten, the common term for a family of digestion resistant, proline and glutamine-rich seed storage proteins found in the cereals wheat, barley and rye. The only current therapy for CD and NCGS is the maintenance of a life-long gluten free diet. Here, I present a brief review of these diseases as well as the role that study of the gut microbiome in humans and a non-human primate (NHP) model of CD can play in identifying treatments to address these conditions. Additionally, I present a brief overview of my research to develop a non-GMO (genetically modified organism), decreased gluten wheat and its relevance and potential applications in the context of CD, NCGS and human nutrition.

#### INTRODUCTION

The most well-known symptoms due to gluten ingestion in coeliac disease sufferers, including diarrhoea and stomach upset, were already described in antiquity about 2000 years ago: these disease manifestations were noted by the Greek physician Aretaeus of Cappadocia in the second century BCE (cited in: (*Freeman*, 2015). A derivation of the term he used "koiliakos", meaning "suffering in the bowels" gave coeliac disease its name (*Fasano* and *Flaherty*, 2014).

Remarkably, however, it wasn't until the middle of the 20<sup>th</sup> century that the definitive association between wheat consumption and the disease was established. In the 1930's the Dutch paediatrician, Dr. Willem-Karel Dicke, formulated the hypothesis that cereals

such as wheat, rye, barley and (sometimes) oats were harmful to his paediatric coeliac patients. However, it was only during, and in the aftermath of the 2<sup>nd</sup> World War that he was able to definitively establish the connection between cereal consumption and deleterious symptoms in his patients. He noted that some of his patients improved when wheat flour was not available due to the deprivations of the war and that their condition worsened again after the war when wheat once again became available and his patients resumed eating wheat as a component of their diets. This, and subsequent careful studies of faecal fat content in his patients, led Dr. Dicke to publish the now generally accepted view that these cereals are the causative factor of coeliac disease (van

Berge-Henegouwen and Mulder, 1993).

In the years since this groundbreaking work there has been an increasing interest in coeliac disease as clinicians gradually became aware that the condition is not as rare as once thought, and as patient groups lobbied for new treatments. Coeliac disease was once thought to be primarily a paediatric condition, but it is now known that it can arise at any age. It is an autoimmune response triggered by the consumption of gluten in genetically susceptible individuals: principally individuals expressing human leukocyte antigen (HLA) haplotypes DQ2 and DQ8 (for a recent review see: Lindfors et al., 2019). In susceptible individuals, the consequence of consumption of wheat, barley, or rye is a chronic intestinal inflammation and flattening of the nutrient-absorbing villi of the small intestine. The clinical presentation of the disease, however, can vary drastically from very severe to no symptoms, to extra-intestinal symptoms such as dermatitis herpetiformis (skin rash) or neurological symptoms (Jackson et al., 2012). Because its symptoms vary so widely, there can oftentimes be a long interval between initial presentation and diagnosis, and the disease is still under-reported.

Nowadays, the disease is identified by screening for auto-antibodies such transglutaminase 2 antibodies (TG2As), and endomysial antibodies (EmAs), as well as antibodies to gliadin. Definitive diagnosis follows with a small intestinal mucosal biopsy while the patient is still on a glutencontaining diet to confirm villous flattening. Once diagnosed, the only treatment is strict adherence to a lifelong gluten free diet. This can be quite challenging since wheat gluten is present in many processed foods, and it imposes an economic burden because most gluten-free substitutes

normally gluten-containing products are over 200% more expensive than their gluten-containing counterparts (*Lee* et al., 2007).

In recent years a number of drug candidates have emerged that are undergoing testing as potential treatments for coeliac disease. These include a modulator of intestinal tight junction permeability (Kelly et al., 2013), a polymer that sequesters gluten (Alhassan et al., 2019), and several competing gluten-degrading enzymes (Sollid and Khosla, 2011; Wolf et al., 2015; Wungjiranirun et al., 2016). There were also high hopes for a therapeutic vaccine (Nexvax2) under development by the company ImmusanT that was in Phase II trials (Di Sabatino et al., 2018). These hopes, however, were dashed with the very recent announcement (June 25, 2019) that ImmusanT was suspending the Phase II trial because interim analysis of the data showed that the vaccine was no better than placebo in providing protection from gluten exposure to patients (http://www.immusant.com/ImmusanT %20Nexvax2%20P2%20-%2025Jun19%20Final.pdf).

With the exception of the now discontinued Nexvax2, the CD therapies in development are not intended to allow CD sufferers to resume an unrestricted diet, rather they are meant to provide protection to patients from small amounts of inadvertently consumed gluten while they are following a gluten free diet.

In addition to coeliac disease, and wheat allergy (not discussed here), a recently identified condition known as non-coeliac gluten sensitivity (NCGS) has emerged. The key similarity with CD is that symptoms in patients resolve with a gluten free diet, but this syndrome is not associated with villous flattening, and the lack of clear biomarkers of disease makes this a still

mysterious condition (*Dale* et al., 2019). Besides gluten, there is evidence that so-called FODMAPS (fermentable oligo-, di-, monosaccharides and polyols) are responsible for symptoms in some NCGS patients. This has led *Dale* et al., (2019) to suggest that a more accurate term for the condition might be non-coeliac wheat sensitivity (NCWS) rather than NCGS, to acknowledge the role that other wheat constituents play in this syndrome, and it might also be

considered a subtype of irritable bowel syndrome (IBS). The prevalence of NCGS is unclear, as many individuals are self-diagnosed and avoid gluten and wheat products out of a belief that such a diet is healthier. Estimates range widely, from less than 1% to greater than 10% of the population (*Aziz* et al., 2016). Until specific biomarkers and other diagnostic criteria are developed, this syndrome will remain somewhat undefined.

## COELIAC DISEASE AND THE MICROBIOME

Considering that the study of the human microbiome is a young field, which was stimulated by the development of the inexpensive "next generation" sequencing technologies that arose out of technology developed for the sequencing of the human genome, the first human microbiome studies were, in effect, exercises in charting the territory and establishing a catalogue of the existing microbial diversity living in the diverse ecological niches present on and in healthy human bodies (*Human Microbiome Project Consortium*, 2012).

Likewise, these are still early days in the study of the human gut microbiome in relation to coeliac disease, and the studies reported to date might be regarded as being in the "cataloguing" phase of identifying differences in the gut microbiomes of coeliac patients compared to matched controls (Sanz et al., 2011). Such studies provide a valuable service in beginning to unravel the interactions between the intestinal pathology of coeliac disease and the attendant gut microbial community. The composition of the intestinal microbiome in effect may serve as a surrogate biomarker of the health of the individual. In addition, changes in the microbiome over time, for example in response to dietary or other interventions, may

serve to document the effect of those interventions (*Lindfors* et al., 2019). Whether identified differences in the microbiomes of coeliac patients compared to healthy controls are a cause or consequence of the disease is not completely clear. Nevertheless, there is evidence that the HLA alleles, which predispose the individuals carrying them to develop coeliac disease, may influence their carriers to develop a disease-predisposing microbiome composition (*Olivares* et al., 2015, 2018; *Sanz*, 2015).

In general, the studies reported to date frequently show microbiome differences between CD individuals and controls but are somewhat inconclusive with regard to functional consequences (Cheng et al., 2013). In many cases, a decreased diversity in the gut microbiomes of coeliac patients compared to healthy controls has been found, frequently with differences in the types of gluten-degrading species between the two groups (Caminero et al., 2014, 2016). In addition, gut dysbiosis in patients compared to healthy subjects is associated with a decrease in the ratio of putatively harmless bacteria (Lactobacillus Bifidobacterium) compared to potenharmful bacteria teroides/Prevotella – E. coli) (Sanz et al., 2011; Sánchez et al., 2013).

Interestingly, one study found that early antibiotic use, which affects the gut microbiome, was associated with the development of coeliac disease (Mårild et al., 2013), however, a later study did not find such an association (Kemppainen et al., 2017). A recent review of the landscape of environmental and genetic factors that influence the development of coeliac disease highlights the role of microbiome composition as a causative factor in the loss of tolerance to gluten in CD patients (Serena et al., 2019).

In our own studies testing the effects of different animal feeds in a non-human primate (rhesus macaque) model of coeliac disease, we found that the faecal microbiome of gluten sensitive animals on a gluten containing diet displayed an overabundance of Strepto-coccaceae and Lactobacillaceae and a

depletion of Coriobacteriaceae compared to healthy controls. In addition, the GS animals exhibited an upregulation of pro-inflammatory miRNAs (Mohan et al., 2016). This is the first assessment of faecal microbiomes in gluten sensitive rhesus macaques and the results strengthen the case for the use of this NHP model of CD to test experimental therapies (Bethune et al., 2008). Experiments to assess the effects on the faecal microbiomes in the NHP model of other dietary interventions, such as decreased gluten wheat, in combination with gluten degrading enzymes, are warranted. Our initial studies in the NHP model using a decreased gluten barley (lys3a mutant) animal feed pave the way for follow-up studies with decreased gluten wheat (Sestak et al., 2015; Sestak et al., 2016).

# CEREAL SEED PROTEIN MUTATIONS: RELEVANCE TO WHEAT GLUTEN, NUTRITION AND CD

The seeds of grasses like rice, wheat and maize directly and indirectly provide most of the calories that feed humanity, thus the mechanisms that control the accumulation of the major storcompounds in seeds, namely starch, proteins and oils, have been of longstanding interest to scientists. In addition, because the storage proteins of many cereal crops such as maize and wheat are deficient in amino acids essential in the human diet and the diets of monogastric animals, considerable efforts have been devoted to identifying genetic variants that increase the amounts of these amino acids, principally lysine, threonine, tryptophan and methionine, in the seeds of cereal crops (Azevedo and Arruda, 2010). One of the earliest known genetic variants whose effect is to increase essential

amino acids in cereal seeds is the opaque2 (02) mutation in maize (Mertz et al., 1964), which leads to reductions in the accumulation of the 22 kilodalton size class of zein seed storage proteins (known as the  $\alpha$  zeins) as well as to increased lysine content in maize seeds. When the gene underlying this mutation was finally isolated over 50 years after the mutant was first identified, it was found to encode a bZIP (basic leucine zipper) transcription factor (TF) that activates transcription of seed storage protein genes by binding to an upstream element found in the promoters of many of these genes (Schmidt et al., 1987) (Schmidt et al., 1990; 1992). Subsequently this mutation, together with genetic modifiers that ameliorate its negative pleiotropic effects, formed the basis for the development of QPM (quality protein maize) varieties (*Prasanna* et al., 2001) containing increased lysine with demonstrated nutritional benefits in humans (*Gunaratna* et al., 2010; *Nuss* and *Tanumihardjo*, 2011).

Since the identification of the transcription factor underlying the o2 mutation, which contains a basic DNAbinding region followed by a proteinprotein interaction motif known as a leucine zipper (Schmidt et al., 1990), multiple additional transcription factors belonging to different structural classes have been determined to play roles regulating processes of seed nutritional reserve accumulation in maize and other cereals. These include transcription factors belonging to the DOF (DNAbinding one zinc finger) family, which interact with the O2-family transcription factors in binding to the promoters of seed storage protein genes (Vicente-Carbajosa et al., 1997; Mena et al., 1998; Zhang et al., 2015).

The success of increasing the content of essential amino acids such as lysine in o2 maize motivated the search for similar mutants in related cereals. A program of barley mutagenesis and screening for increased lysine using a dye binding technique at the Risø agricultural experiment station in Roskilde, Denmark in the 1960s led to the identification of numerous enhanced lysine mutants of which the best characterized is the lys3a mutant (Køie and Doll, 1979; Miflin and Shewry, 1979). This mutant is also referred to as Risø 1508 or sex3c (shrunken endosperm xenia) (Ullrich and Eslick, 1977); it was induced by ethylenimine mutagenesis of the two-rowed spring malting variety, Bomi (Doll et al., 1974). The lys3a mutant is almost completely lacking in class C hordeins, and accumulates considerably reduced amounts of several B class hordeins, while having a 45% increase in the accumulation of free and

protein bound lysine in the seed compared to the parental variety (Shewry et al., 1977; Shewry et al., 1978; Munck et al., 2001). These phenotypes are the result of a single recessive allele (*Doll*, 1973) that was mapped to barley chromosome 5H (Karlsson, 1977; Ullrich and Eslick, 1977; Jensen, 1979), although more recent research asserts that this map position is erroneous and that the mutant is located on chromosome 1H (Druka et al., 2011; Rustgi and von Wettstein, 2015). The underlying lesion responsible for the *lys3a* mutant's pleiotropic effects was hypothesized to be in a 5-methylcytosine DNA glycosylase gene (DEMETER, DNA demethylase) (Gehring et al., 2009) responsible for removing the methyl groups from methylated DNA since it was found that the methylation state of the promoters of several seed storage protein genes was altered in the mutant (Sørensen, 1992; Sørensen et al., 1996) (Wen et al., 2012).

The original *lys3a* mutant suffered from negative pleiotropic effects including lower yield and shrunken seeds containing reduced total starch. Nevertheless, concerted breeding efforts minimized these effects (Eggum et al., 1995; Jørgensen et al., 1999). Barley varieties containing the *lys3a* allele, however, were still lower in starch content than conventional cultivars and for this reason they were never widely grown (*Munck* and *Jespersen*, 2009). In recent years, there has been a resurgence of interest in this mutant, not because of its higher lysine content, but due to its lower levels of several classes of hordeins. The *lys3a* mutant, in combination with several other mutations that reduce the accumulation of hordeins, has been used to create ultralow gluten barley (ULG) (*Tanner* et al., 2016). In addition, we tested the *lys3a* mutant in the rhesus macaque model of coeliac disease to determine if it, alone or in combination with a gluten-degrading enzyme supplement, can ameliorate the symptoms of gluten sensitivity in this NHP model of coeliac disease (*Sestak* et al., 2015; 2016). In our 2016 paper, we reported that a diet of this *lys3a* mutant barley feed, combined with a gluten-degrading enzyme (Tolerase G, manufactured by DSM), appeared to eliminate symptoms due to gluten sensitivity in this animal model of coeliac disease.

We subsequently determined that a missense mutation in the Barley Prolamin-box Binding Factor (BPBF) represents the molecular lesion underlying the *lys3a* mutation (*Moehs* et al., 2019). This barley transcription factor is homologous to a domain of one finger (DOF) zinc finger transcription factor found in maize that has been shown to interact with O2 to control the expression of maize zein seed storage proteins (*Zhang* et al., 2015). A wheat homolog of BPBF had also been identified (*Ravel* et al., 2006).

This allowed us to use the nontransgenic, reverse genetic technique known as TILLING (Targeting Induced Local Lesions in Genomes; Colbert et al., 2001), a variation on mutation breeding, to identify 488 novel wheat lines that contain induced variants in the A, B and D genome copies of the WPBF genes in hexaploid bread wheat. Combining inactivating (recessive) alleles in all three homoeologous copies of the wheat PBF gene in hexaploid bread wheat led to a reduction in the accumulation of several classes of wheat gliadins and low molecular weight glutenins and to an increase in the accumulation of free and proteinbound lysine in wheat endosperm. These lines, which we refer to as decreased gluten wheat (DGW), have lower amounts of the epitopes that are detrimental to coeliac patients, but they are not low enough in gluten to be safe to consume for CD patients. However, they are novel in that they contain levels of gluten that are lower and outside of the range of the available genetic variation in existing wheat germplasm, including such "ancient" wheats as einkorn and spelt. In addition, there are ongoing efforts to decrease the gluten even further.

At present, the alleles are being introgressed into elite varieties and functional tests are being conducted to assess the types of products (cookies, cake, bread, noodles, tortillas, etc.) for which the DGW is best suited. Based on the outcomes of these tests, combined with other factors, will influence which market class(es) of wheat the alleles will be introgressed into. There are at least 6 market classes of wheat, including spring and winter wheats, as well as hard red and white wheats, in addition to durum pasta wheat. These different market classes all have particular functionality and uses.

A number of other groups are pursuing the development of decreased gluten wheat by other methods including RNAi (Gil-Humanes et al., 2010), and CRISPR (Sánchez-León et al., 2017). These methods have been very effective at creating transcriptional shutdowns in the expression of gliadins in the case of RNAi, or gene deletions of gliadins in the case of CRISPR, but these wheats face additional hurdles that come with being classified as transgenic and thus will face additional public scrutiny and a costlier path to market (Laursen 2016; Jouanin et al., 2018; Hundleby and Harwood, 2019).

#### **CONCLUSION**

The incidence of coeliac disease and non-coeliac gluten (wheat) sensitivity has been on a slow upward trajectory in recent decades and this is not simply due to increased testing and awareness (Rubio-Tapia and Murray, 2010). During the Swedish paediatric coeliac disease epidemic of the 1980s and 1990s, for example, the incidence reached about 3% of tested 12- year-old children, well above previously known rates of the disease (Myléus et al., 2009). Given that the predisposing HLA alleles are quite common in most human populations, but only a small subset of individuals carrying these alleles lose tolerance to gluten and develop coeliac disease, considerable effort has been devoted to identifying possible environmental variables that may contribute to loss of gluten tolerance (reviewed in: Lindfors et al., 2019)). Among the possible identified factors include infections with intestinal viruses (*Bouziat* et al., 2017; *Kahrs* et al., 2019), and a recent study suggested that vaccination against rotavirus may decrease paediatric coeliac disease cases (Hemming-Harlo et al., 2019). The seasonality of intestinal virus infections in children may also play a role in the link identified between seasonality of birth and the development of coeliac disease (Daniel et al., 2019). In the case of the Swedish CD epidemic, it was proposed that changes in the amount and timing of the introduction of wheat into infants' diets, as well as whether wheat was introduced while infants were still breastfeeding, played a role in the subsequent development of coeliac disease (Ivarsson et al., 2002). A recent meta-analysis, however, did not support a role for breastfeeding in subsequent development of CD (Szajewska et al., 2015) nor did adjusting the timing of gluten introduction into the diet of genetically predisposed infants prevent the development of CD in a well-designed, double-blind, randomized, placebocontrolled intervention study (*Vriezinga* et al., 2014).

Against the backdrop of clinical disease there is also a cultural shift occurring in many western nations with a dramatic increase in, and availability of, gluten free products, the selfdiagnosis of "gluten sensitivity" and adoption of a gluten free and/or a low carbohydrate diet by a significant fraction of the public even in the absence of disease. This has been stimulated in part by a spate of best-selling, nonscientific books with sensationalistic titles such as "Wheat belly: lose the wheat, lose the weight, and find your path back to health," and "Grain brain: the surprising truth about wheat, carbs, and sugar--your brain's silent killers" that hold the consumption of wheat responsible for a host of current western societal ills, from "brain fog", to fatigue to obesity (Davis, 2014; Perlmutter and Loberg, 2018). In the current climate, it is not surprising that sensationalistic claims received more notice than the sober, scientific rebuttals (Jones, 2012; Brouns et al., 2013).

The "DIY" approach to gluten sensitive self-diagnosis even in the absence of a clinical diagnosis has also laudable aspects in the sense of the desire to control one's own health. One example of this is the growth in genetic self-testing companies such as 23 and Me and its rivals. Self-testing of one's own microbiome is not far behind; there are already companies that offer this testing as a service, and individual accounts of dietary self-experiments and effects on the individual's own microbiome have been published (*Sprague*, 2017). Although there is no

"microbiome pill" to cure CD or NCGS, a number of microbiome companies (Second Genome, LNC Therapeutics) are dissecting the human microbiome with the objective to develop probiotics or pharmaceutical compounds to modulate disease states such as inflammatory bowel disease (IBD). Nature Publishing Group has just published a special collection of papers from the NIH funded Human Microbiome **Project** (https://www.nature.com/collections/fia bfcjbfj), and this is a research area that will likely continue to produce exciting insights and innovations in human health care in the years ahead (*Proctor*, 2019).

The decreased hordein (lys3a) barley that we used as a model in our studies during the course of the development of decreased gluten wheat appeared to positively impact the gut microbiomes of gluten sensitive rhesus macaques (Mohan et al., 2016), and it is to be hoped that similar follow-up studies will be conducted with decreased gluten wheat in the future. Along these lines, a recent report claims that a diet of the ancient diploid wheat, einkorn, induced beneficial changes in the gut microbiomes of pigs compared to a diet of conventional wheat (*Barone* et al., 2018), including increased abundance of the putatively health-associated species Oscillospira (Konikoff and Gophna, 2016). These microbiome changes, however, are unlikely to be related to the (minor) differences in the gluten composition between einkorn and conventional wheat used in the study, but more likely to be related to other compositional differences between these different wheats.

It may be that the increased awareness of gluten sensitivity in the public consciousness, an interest in low carbohydrate diets, as well as a desire to purchase local, organically grown food, has contributed to the rapid recent rise in the demand for and marketing of the so-called ancient wheats: einkorn, hulled spelt, and Kamut®. Although it has not been substantiated that these still niche types of wheat are lower in gluten and thus healthier for individuals with gluten sensitivity (Dinu et al., 2018), their market rise is a signal of consumer demand. Any decreased gluten wheat that ultimately comes to market might follow a similar market trajectory. The story of Quality Protein Maize, likewise, offers an encouraging parallel with DGW, in that QPM ultimately emerged as an agronomically and functionally beneficial variety with proven health benefits. While DGW must still surmount agronomic, functional and health and nutritional tests before it has any impact, the field ahead is full of promise.

#### **LITERATURE**

Alhassan, E., Yadav, A., Kelly, C.P., and Mukherjee, R.: Novel nondietary therapies for celiac disease. Cell. Mol. Gastroenterol. Hepatol. 8, 335-345 (2019).

Azevedo, R. and Arruda, P.: High-lysine maize: The key discoveries that have made it possible. Amino acids 39, 979-989 (2010).

Aziz, I., Dwivedi, K., and Sanders, D.S.: From

coeliac disease to noncoeliac gluten sensitivity; should everyone be gluten free? Curr. Opin. Gastroenterol. 32, 120-127 (2016).

Barone, F., Laghi, L., Gianotti, A., Ventrella, D., Saa, D.L.T., Bordoni, A., Forni, M., Brigidi, P., Bacci, M.L., and Turroni, S.: In vivo effects of einkorn wheat (*Triticum monococcum*) bread on the intestinal

- microbiota, metabolome, and on the glycemic and insulinemic response in the pig model. Nutrients 11, E16 (2018).
- Bethune, M.T., Borda, J.T., Ribka, E., Liu, M.X., Phillippi-Falkenstein, K., Jandacek, R.J., Doxiadis, G.G.M., Gray, G.M., Khosla, C., and Sestak, K.: A non-human primate model for gluten sensitivity. PLoS One 3, e1614 (2008).
- Bouziat, R., Hinterleitner, R., Brown, J.J.,
  Stencel-Baerenwald, J.E., Ikizler, M., Mayassi, T., Meisel, M., Kim, S.M., Discepolo, V., Pruijssers, A.J., Ernest, J.D., Iskarpatyoti, J.A., Costes, L.M., Lawrence, I., Palanski, B.A., Varma, M., Zurenski, M.A., Khomandiak, S., McAllister, N., Aravamudhan, P., Boehme, K.W., Hu, F., Samsom, J.N., Reinecker, H.C., Kupfer, S.S., Guandalini, S., Semrad, C.E., Abadie, V., Khosla, C., Barreiro, L.B., Xavier, R.J., Ng, A., Dermody, T.S., and Jabri, B.: Reovirus infection triggers inflammatory responses to dietary antigens and development of celiac disease. Science 356, 44-50 (2017).
- Brouns, F.J., van Buul, V.J., and Shewry, P.R.: Does wheat make us fat and sick? J. Cereal Sci. 58, 209-215 (2013).
- Caminero, A., Herrán, A.R., Nistal, E., Pérez-Andrés, J., Vaquero, L., Vivas, S., Ruiz de Morales, J.M.G., Albillos, S.M., and Casqueiro, J.: Diversity of the cultivable human gut microbiome involved in gluten metabolism: Isolation of microorganisms with potential interest for coeliac disease. FEMS Microbiol. Ecol. 88, 309-319 (2014).
- Caminero, A., Galipeau, H.J., McCarville, J.L., Johnston, C.W., Bernier, S.P., Russell, A.K., Jury, J., Herran, A.R., Casqueiro, J., Tye-Din, J.A., Surette, M.G., Magarvey, N.A., Schuppan, D., and Verdu, E.F.: Duodenal bacteria from patients with celiac disease and healthy subjects distinctly affect gluten breakdown and immunogenicity. Gastroenterology 151, 670-683 (2016).
- Cheng, J., Kalliomäki, M., Heilig, H.G., Palva, A., Lähteenoja, H., de Vos, W.M., Salojärvi, J., and Satokari, R.: Duodenal microbiota composition and mucosal homeostasis in pediatric celiac disease.

- BMC Gastroenterol. 13, 113 (2013).
- Colbert, T., Till, B.J., Tompa, R., Reynolds, S., Steine, M.N., Yeung, A.T., McCallum, C.M., Comai, L., Henikoff, S.: Highthroughput screening for induced point mutations. Plant Physiol. 126, 480-484 (2001).
- Dale, H.F., Biesiekierski, J.R., and Lied, G.A.: Non-coeliac gluten sensitivity and the spectrum of gluten-related disorders: An updated overview. Nutr. Res. Rev. 32, 28-37 (2019).
- Daniel, S., Kalansky, A., Tsur, A., Pinsk, V., Ling, G., Rannan, R., and Yerushalmi, B.: Seasonality of birth affects paediatric coeliac disease. Acta Paediatr. 108, 529-534 (2019).
- Davis, W.: Wheat belly: Lose the wheat, lose the weight, and find your path back to health. Rodale Books, Emmaus, Pensylvania (2014).
- Di Sabatino, A., Lenti, M.V., Corazza, G.R., and Gianfrani, C.: Vaccine immunotherapy for celiac disease. Front. Med. (Lausanne) 5, 187 (2018).
- Dinu, M., Whittaker, A., Pagliai, G., Benedettelli, S., and Sofi, F.: Ancient wheat species and human health: Biochemical and clinical implications. J. Nutr. Biochem. 52, 1-9 (2018).
- Doll, H.: Inheritance of the high-lysine character of a barley mutant. Hereditas 74, 293-294 (1973).
- Doll, H., Køie, B., and Eggum, B.O.: Induced high lysine mutants in barley. Radiat. Botany 14, 73-80 (1974).
- Druka, A., Franckowiak, J., Lundqvist, U., Bonar, N., Alexander, J., Houston, K., Radovic, S., Shahinnia, F., Vendramin, V., Morgante, M., Stein, N., and Waugh, R.: Genetic dissection of barley morphology and development. Plant Physiol. 155, 617-627 (2011).
- Eggum, B.O., Brunsgaard, G., and Jensen, J.: The nutritive value of new high-lysine barley mutants. J. Cereal Sci. 22, 171-176 (1995).
- Fasano, A. and Flaherty, S.: Gluten Freedom. Turner Publishing Company, Nashville, Tennessee (2014).

- Freeman, H.J.: Celiac disease: A disorder emerging from antiquity, its evolving classification and risk, and potential new treatment paradigms. Gut Liver 9, 28-37 (2015).
- Gehring, M., Reik, W., and Henikoff, S.: DNA demethylation by DNA repair. Trends Genet. 25, 82-90 (2009).
- Gil-Humanes, J., Pistón, F., Tollefsen, S., Sollid, L.M., and Barro, F.: Effective shutdown in the expression of celiac disease-related wheat gliadin T-cell epitopes by RNA interference. Proc. Natl. Acad. Sci. USA 107, 17023-17028 (2010).
- Gunaratna, N.S., Groote, H.D., Nestel, P., Pixley, K.V., and McCabe, G.P.: A meta-analysis of community-based studies on quality protein maize. Food Policy 35, 202-210 (2010).
- Hemming-Harlo, M., Lähdeaho, M.L., Mäki, M., and Vesikari, T.: Rotavirus vaccination does not increase type 1 diabetes and may decrease celiac disease in children and adolescents. Pediatr. Infect. Dis. J. 38, 539-541 (2019).
- Human Microbiome Project Consortium: Huttenhower, C., Gevers, D., Knight, R., Abubucker, S., Badger, J.H., Chinwalla, A.T., Creasy, H.H., Earl, A.M., FitzGerald, M.G., Fulton, R.S., Giglio, M.G., Hallsworth-Pepin, K., Lobos, E.A., Madupu, R., Magrini, V., Martin, J.C., Mitreva, M., Muzny, D.M., Sodergren, E.J., Versalovic, J., Wollam, A.M., Worley, K.C., Wortman, J.R., Young, S.K., Zeng, Q., Aagaard, K.M., Abolude, O.O., Allen-Vercoe, E., Alm, E.J., Alvarado, L., Andersen, G.L., Anderson, S., Appelbaum, E., Arachchi, H.M., Armitage, G., Arze, C.A., Ayvaz, T., Baker, C.C., Begg, L., Belachew, T., Bhonagiri, V., Bihan, M., Blaser, M.J., Bloom, T., Bonazzi. V., Brooks, J., Buck, G.A., Buhay, C.J., Busam, D.A., Campbell, J.L., Canon, S.R., Cantarel, B.L., Chain, P.S., Chen, I.M., Chen, L., Chhibba, S., Chu, K., Ciulla, D.M., Clemente, J.C., Clifton, S.W., Conlan, S., Crabtree, J., Cutting, M.A., Davidovics, N.J., Davis, C.C., DeSantis, T.Z., Deal, C., Delehaunty, K.D., Dewhirst, F.E., Deych, E., Ding, Y., Dooling, D.J., Dugan,

S.P., Dunne, W.M., Durkin, A., Edgar, R.C., Erlich, R.L., Farmer, C.N., Farrell, R.M., Faust, K., Feldgarden, M., Felix, V.M., Fisher, S., Fodor, A.A., Forney, L.J., Foster, L., Di Francesco, V., Friedman, J., Friedrich, D.C., Fronick, C.C., Fulton, L.L., Gao H., Garcia, N., Giannoukos, G., Giblin, C., Giovanni, M.Y., Goldberg, J.M., Goll, J., Gonzalez, A., Griggs, A., Gujja, S., Haake, S.K., Haas, B.J., Hamilton, H.A., Harris, E.L., Hepburn, T.A., Herter, B., Hoffmann, D.E., Holder, M.E., Howarth, C., Huang, K.H., Huse, S.M., Izard, J., Jansson, J.K., Jiang, H., Jordan, C., Joshi, V., Katancik, J.A., Keitel, W.A., Kelley, S.T., Kells, C., King, N.B., Knights, D., Kong, H.H., Koren, O., Koren, S., Kota, K.C., Kovar, C.L., Kyrpides, N.C., La Rosa, P.S., Lee, S.L., Lemon, K.P., Lennon, N., Lewis, C.M., Lewis, L., Ley, R.E., Li, K., Liolios, K., Liu, B., Liu, Y., Lo, C.C., Lozupone, C.A., Lunsford, R., Madden, T., Mahurkar, A.A., Mannon, P.J., Mardis, E.R., Markowitz, V.M., Mavromatis, K., McCorrison, J.M., McDonald, D., McEwen, J., McGuire, A.L., McInnes, P., Mehta, T., Mihindukulasuriya, K.A., Miller, J.R., Minx, P.J., Newsham, I., Nusbaum, C., O'Laughlin, M., Orvis, J., Pagani, I., Palaniappan, K., Patel, S.M., Pearson, M., Peterson, J., Podar, M., Pohl, C., Pollard, K.S., Pop, M., Priest, M.E., Proctor, L.M., Qin, X., Raes, J., Ravel, J., Reid, J.G., Rho, M., Rhodes, R., Riehle, K.P., Rivera, M.C., Rodriguez-Mueller, B., Rogers, Y.H., Ross, M.C., Russ, C., Sanka, R.K., Sankar, P., Sathirapongsasuti, J., Schloss, J.A., Schloss, P.D., Schmidt, T.M., Scholz, M., Schriml, L., Schubert, A.M., Segata, N., Segre, J.A., Shannon, W.D., Sharp, R.R., Sharpton, T.J., Shenoy, N., Sheth, N.U., Simone, G.A., Singh, I., Smillie, C.S., Sobel, J.D., Sommer, D.D., Spicer, P., Sutton, G.G., Sykes, S.M., Tabbaa, D.G., Thiagarajan, M., Tomlinson, C.M., Torralba, M., Treangen, T.J., Truty, R.M., Vishnivetskaya, T.A., Walker, J., Wang, L., Wang, Z., Ward, D.V., Warren, W., Watson, M.A., Wellington, C., Wetterstrand, K.A., White, J.R., Wilczek-

- Boney, K., Wu, Y., Wylie, K.M., Wylie, T., Yandava, C., Ye, L., Ye, Y., Yooseph, S., Youmans, B.P., Zhang, L., Zhou, Y., Zhu, Y., Zoloth, L., Zucker, J.D., Birren, B.W., Gibbs, R.A., Highlander, S.K., Methé, B.A., Nelson, K.E., Petrosino, J.F., Weinstock, G.M., Wilson, R.K., and White, O.: Structure, function and diversity of the healthy human microbiome. Nature 486, 207-214 (2012).
- Hundleby, P.A.C. and Harwood, W.A.: Impacts of the EU GMO regulatory framework for plant genome editing. Food Energy Secur. 8, e00161 (2019).
- Ivarsson, A., Hernell, O., Stenlund, H., and Persson, L.Å.: Breast-feeding protects against celiac disease. Am. J. Clin. Nutr. 75, 914-921 (2002).
- Jackson, J.R., Eaton, W.W., Cascella, N.G., Fasano. A., and Kelly. D.L.: Neurologic and psychiatric manifestations of celiac disease and gluten sensitivity. Psychiatr. Q. 83, 91-102 (2012).
- Jensen, J.: Location of a high-lysine gene and the DDT-resistance gene on barley chromosome 7. Euphytica 28, 47-56 (1979).
- Jones, J.: Wheat belly—an analysis of selected statements and basic theses from the book. Cereal Foods World 57, 177-189 (2012).
- Jørgensen, H., Gabert, V.M., and Fernández, J.A.: Influence of nitrogen fertilization on the nutritional value of high-lysine barley determined in growing pigs. Anim. Feed Sci. Technol. 79, 79-91 (1999).
- Jouanin, A., Boyd, L.A., Visser, R.G.F., and Smulders, M.J.: Development of wheat with hypoimmunogenic gluten obstructed by the gene editing policy in Europe. Front. Plant Sci. 9, 1523 (2018).
- Kahrs, C.R., Chuda, K., Tapia, G., Stene, L.C., Mårild, K., Rasmussen, T., Rønningen, K.S., Lundin, K.E., Kramna, L., and Cinek, O.: Enterovirus as trigger of coeliac disease: Nested case-control study within prospective birth cohort. BMJ 364, 1231 (2019).
- Karlsson, K.: Linkage studies in a gene for high lysine content in Riso barley mutant 1508. Barley Genet. Newsl. 7, 40-43 (1977).

- Kelly, C., Green, P., Murray, J.A., Dimarino, A., Colatrella, A., Leffler, D., Alexander, T., Arsenescu, R., Leon, F., and Jiang, J.: Larazotide acetate in patients with coeliac disease undergoing a gluten challenge: A randomised placebo-controlled study. Aliment. Pharmacol. Ther. 37, 252-262 (2013).
- Kemppainen, K.M., Vehik, K., Lynch, K.F., Larsson, H.E., Canepa, R.J., Simell, V., Koletzko, S., Liu, E., Simell, O.G., Toppari, J. Ziegler, A.G., Rewers, M.J., Lernmark, Å., Hagopian, W.A., She, J.X., Akolkar, B., Schatz, D.A., Atkinson, M.A., Blaser, M.J., Krischer, J.P., Hyöty, H., Agardh, D., and Triplett, E.W.; Environmental determinants of diabetes in the young (TEDDY) study group: Association between early-life antibiotic use and the risk of islet or celiac disease autoimmunity. JAMA Pediatr. 171, 1217-1225 (2017).
- Køie, B. and Doll, H.: Protein and carbohydrate components in the Risø high-lysine barley mutants. In: Seed protein improvement in cereals and grain legumes, Vol. 1. International Atomic Energy Agency (IAEA), Vienna, 205-214 (1979).
- Konikoff, T. and Gophna, U.: Oscillospira: A central, enigmatic component of the human gut microbiota. Trends Microbiol. 24, 523-524 (2016).
- Laursen, L.: Will Europe toast GM wheat for gluten sufferers? Nat. Biotechnol. 34, 369-371 (2016).
- Lee, A.R., Ng, D.L., Zivin, J., and Green, P.H.R.: Economic burden of a gluten-free diet. J. Hum. Nutr. Diet. 20, 423-430 (2007).
- Lindfors, K., Ciacci, C., Kurppa, K., Lundin, K.E., Makharia, G.K., Mearin, M.L., Murray, J.A., Verdu, E.F., and Kaukinen, K.: Coeliac disease. Nat. Rev. Dis. Primers 5, 3 (2019).
- Mårild, K., Ye, W., Lebwohl, B., Green, P.H., Blaser, M.J., Card, T., and Ludvigsson, J.F.: Antibiotic exposure and the development of coeliac disease: A nationwide case-control study. BMC Gastroenterol. 13, 109 (2013).
- Mena, M., Vicente-Carbajosa, J., Schmidt, R.J., and Carbonero, P.: An endosperm-specific

- DOF protein from barley, highly conserved in wheat, binds to and activates transcription from the prolamin-box of a native Bhordein promoter in barley endosperm. Plant J. 16, 53-62 (1998).
- Mertz, E.T., Bates, L.S., and Nelson, O.E.: Mutant gene that changes protein composition and increases lysine content of maize endosperm. Science 145, 279-280 (1964).
- Miflin, B. and Shewry, P.: The synthesis of proteins in normal and high lysine barley seeds. In: Recent advances in the biochemistry of cereals (Annual Proceedings of the Phytochemical Society of Europe) (Eds.: Laidman, D.L. and Wyn Jones, R.G.). Academic Press, London, New York, 239-273 (1979).
- Moehs, C.P., Austill, W.J., Holm, A., Large, T.A., Loeffler, D., Mullenberg, J., Schnable, P.S., Skinner, W., van Boxtel, J., Wu, L., and McGuire, C.: Development of decreased-gluten wheat enabled by determination of the genetic basis of *lys3a* barley. Plant Physiol. 179, 1692-1703 (2019).
- Mohan, M., Chow, C.-E.T., Ryan, C.N., Chan, L.S., Dufour, J., Aye, P.P., Blanchard, J., Moehs, C.P., and Sestak, K.: Dietary gluten-induced gut dysbiosis is accompanied by selective upregulation of microRNAs with intestinal tight junction and bacteriabinding motifs in rhesus macaque model of celiac disease. Nutrients 8, 684 (2016).
- Munck, L., Pram Nielsen, J., Møller, B., Jacobsen, S., Søndergaard, I., Engelsen, S.B., Nørgaard, L., and Bro, R.: Exploring the phenotypic expression of a regulatory proteome-altering gene by spectroscopy and chemometrics. Anal. Chim. Acta 446, 169-184 (2001).
- Munck, L. and Jespersen, B.P.M.: The multiple uses of barley endosperm mutants in plant breeding for quality and for revealing functionality in nutrition and food technology. In: Induced plant mutations in the genomics era (Ed.: Shu, Q.Y.). Food and Agriculture Organization of the United Nations, Rome, 182-186 (2009).
- Myléus, A., Ivarsson, A., Webb, C., Danielsson, L., Hernell, O., Högberg, L.,

- Karlsson, E., Lagerqvist, C., Norström, F., Rosén, A., Sandström, O., Stenhammar, L., Stenlund, H., Wall, S., and Carlsson, A.: Celiac disease revealed in 3% of Swedish 12-year-olds born during an epidemic. J. Pediatr. Gastroenterol. Nutr. 49, 170-176 (2009).
- Nuss, E.T. and Tanumihardjo, S.A.: Quality protein maize for Africa: Closing the protein inadequacy gap in vulnerable populations. Adv. Nutr. 2, 217-224 (2011).
- Olivares, M., Neef, A., Castillejo, G., De Palma, G., Varea, V., Capilla, A., Palau, F., Nova, E., Marcos, A., Polanco, I., Ribes-Koninckx, C., Ortigosa, L., Izquierdo, L., and Sanz, Y.: The HLA-DQ2 genotype selects for early intestinal microbiota composition in infants at high risk of developing coeliac disease. Gut 64, 406-417 (2015).
- Olivares, M., Walker, A.W., Capilla, A., Benítez-Páez, A., Palau, F., Parkhill, J., Castillejo, G., and Sanz, Y.: Gut microbiota trajectory in early life may predict development of celiac disease. Microbiome 6, 36 (2018).
- Perlmutter, D. and Loberg, K.: Grain brain: The surprising truth about wheat, carbs, and sugar--your brain's silent killers. Little, Brown and Company, New York, Boston, London (2018).
- Prasanna, B.M., Vasal, S.K., Kassahun, B., and Singh, N.N.: Quality protein maize. Curr. Sci. 81, 1308-1319 (2001).
- Proctor, L.: Priorities for the next 10 years of human microbiome research. Nature 569, 623-625 (2019).
- Ravel, C., Nagy, I.J., Martre, P., Sourdille, P., Dardevet, M., Balfourier, F., Pont, C., Giancola, S., Praud, S., and Charmet, G.: Single nucleotide polymorphism, genetic mapping, and expression of genes coding for the DOF wheat prolamin-box binding factor. Funct. Integr. Genomics 6, 310-321 (2006).
- Rubio-Tapia, A. and Murray, J.A.: Celiac disease. Curr. Opin. Gastroenterol. 26, 116-122 (2010)
- Rustgi, S. and von Wettstein, D.: Breeding barley ornamented with the novel agronomical

- attributes. Med. Aromat. Plants 4, e158 (2015).
- Sánchez-León, S., Gil-Humanes, J., Ozuna, C.V., Giménez, M.J., Sousa, C., Voytas, D.F., and Barro, F.: Low-gluten, nontransgenic wheat engineered with CRISPR/Cas9. Plant Biotechnol. J. 16, 902-910 (2017).
- Sánchez, E., Donat, E., Ribes-Koninckx, C., Fernández-Murga, M.L., and Sanz, Y.: Duodenal-mucosal bacteria associated with celiac disease in children. Appl. Environ. Microbiol. 79, 5472-5479 (2013)
- Sanz, Y., Palma, G.D., and Laparra, M.: Unraveling the ties between celiac disease and intestinal microbiota. Int. Rev. Immunol. 30, 207-218 (2011).
- Sanz, Y.: Microbiome and gluten. Ann. Nutr. Metab. 67, Suppl. 2, 28-41 (2015).
- Schmidt, R.J., Burr, F.A., and Burr, B.: Transposon tagging and molecular analysis of the maize regulatory locus opaque-2. Science 238, 960-963 (1987).
- Schmidt, R.J., Burr, F.A., Aukerman, M.J., and Burr, B.: Maize regulatory gene opaque-2 encodes a protein with a "leucine-zipper" motif that binds to zein DNA. Proc. Natl. Acad. Sci. USA 87, 46-50 (1990).
- Schmidt, R.J., Ketudat, M., Aukerman, M.J., and Hoschek, G.: Opaque-2 is a transcriptional activator that recognizes a specific target site in 22-kD zein genes. Plant Cell 4, 689-700 (1992).
- Serena, G., Lima, R., and Fasano, A.: Genetic and environmental contributors for celiac disease. Curr. Allergy Asthma Rep. 19, 40 (2019).
- Sestak, K., Thwin, H., Dufour, J., Aye, P.P., Liu, D.X., and Moehs, C.P.: The effects of reduced gluten barley diet on humoral and cell-mediated systemic immune responses of gluten-sensitive rhesus macaques. Nutrients 7, 1657-1671 (2015).
- Sestak, K., Thwin, H., Dufour, J., Liu, D.X., Alvarez, X., Laine, D., Clarke, A., Doyle, A., Aye, P.P., Blanchard, J., and Moehs, C.P.: Supplementation of reduced gluten barley diet with oral prolyl endopeptidase effectively abrogates enteropathy-

- associated changes in gluten-sensitive macaques. Nutrients 8, E401 (2016).
- Shewry, P., Pratt, H.M., Charlton, M., and Miflin, B.: Two-dimensional separation of the prolamins of normal and high lysine barley (*Hordeum vulgare* L.). J. Exp. Bot. 28, 597-606 (1977).
- Shewry, P., Hill, J., Pratt, H., Leggatt, M., and Miflin, B.: An evaluation of techniques for the extraction of hordein and glutelin from barley seed and a comparison of the protein composition of Bomi and Risø 1508. J. Exp. Bot. 29, 677-692 (1978).
- Sollid, L.M. and Khosla, C.: Novel therapies for coeliac disease. J. Intern. Med. 269, 604-613 (2011).
- Sørensen, M.B.: Methylation of B-hordein genes in barley endosperm is inversely correlated with gene activity and affected by the regulatory gene Lys3. Proc. Natl. Acad. Sci. USA 89, 4119-4123 (1992).
- Sørensen, M.B., Muller, M., Skerrit, J., and Simpson, D.: Hordein promoter methylation and transcriptional activity in wild type and mutant barley endosperm. Mol. Gen. Genet. 250, 750-760 (1996).
- Sprague, R.: Fusicatenibacter is associated with kefir drinking. BioRxiv, 218313 (2017).
- Szajewska, H., Shamir, R., Chmielewska, A., Pieścik-Lech, M., Auricchio, R., Ivarsson, A., Kolacek, S., Koletzko, S., Korponay-Szabo, I., and Mearin, M.: Systematic review with meta-analysis: Early infant feeding and coeliac disease--update 2015. Aliment. Pharmacol. Ther. 41, 1038-1054 (2015).
- Tanner, G.J., Blundell, M.J., Colgrave, M.L., and Howitt, C.A.: Creation of the first ultralow gluten barley (*Hordeum vulgare* L.) for coeliac and gluten-intolerant populations. Plant Biotechnol. J. 14, 1139-1150 (2016).
- Ullrich, S.E. and Eslick, R.F.: Inheritance of the shrunken endosperm character, sex3c, of Bomi Riso mutant 1508 and its association with lysine content. Barley Genet. Newsl. 7, 66-73 (1977).
- van Berge-Henegouwen, G. and Mulder, C.: Pioneer in the gluten free diet: Willem-Karel Dicke 1905-1962, over 50 years of

- gluten free diet. Gut 34, 1473-1475 (1993)
- Vicente-Carbajosa, J., Moose, S.P., Parsons, R.L., and Schmidt, R.J.: A maize zinc finger protein binds the prolamin box in zein gene promoters and interacts with the basic leucine zipper transcriptional activator Opaque-2. Proc. Natl. Acad. Sci. USA 94, 7685-7690 (1997).
- Vriezinga, S.L., Auricchio, R., Bravi, E., Castillejo, G., Chmielewska, A., Crespo Escobar, P., Kolaček, S., Koletzko, S., Korponay-Szabo, I.R., Mummert, E., Polanco, I., Putter, H., Ribes-Koninckx, C., Shamir, R., Szajewska, H., Werkstetter, K., Greco, L., Gyimesi, J., Hartman, C., Hogen Esch, C., Hopman, E., Ivarsson, A., Koltai, T., Koning, F., Martinez-Ojinaga, E., te Marvelde, C., Pavic, A., Romanos, J., Stoopman, E., Villanacci, V., Wijmenga, C., Troncone, R., and Mearin, M.L.: Randomized feeding intervention in infants at high risk for celiac disease. N. Engl. J. Med. 371, 1304-1315 (2014).
- Wen, S., Wen, N., Pang, J., Langen, G., Brew-Appiah, R.A., Mejias, J.H., Osorio, C.,

- Yang, M., Gemini, R., Moehs, C.P., Zemetra, R.S., Kogel, K.H., Liu, B., Wang, X., von Wettstein, D., and Rustgi, S.: Structural genes of wheat and barley 5-methylcytosine DNA glycosylases and their potential applications for human health. Proc. Natl. Acad. Sci. USA 109, 20543-20548 (2012).
- Wolf, C., Siegel, J.B., Tinberg, C., Camarca, A., Gianfrani, C., Paski, S., Guan, R., Montelione, G., Baker, D., and Pultz, I.S.: Engineering of Kuma030: A gliadin peptidase that rapidly degrades immunogenic gliadin peptides in gastric conditions. J. Am. Chem. Soc. 137, 13106-13113 (2015).
- Wungjiranirun, M., Kelly, C., and Leffler, D.: Current status of celiac disease drug development. Am. J. Gastroenterol. 111, 779-786 (2016).
- Zhang, Z., Yang, J., and Wu, Y.: Transcriptional regulation of zein gene expression in maize through the additive and synergistic action of opaque2, prolamine-box binding factor, and O2 heterodimerizing proteins. Plant Cell 27, 1162-1172 (2015).