THE HUMAN INTESTINAL MICROBIOTA; FROM PHYLOGENETICS TO FUNCTIONAL METAGENOMICS

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SUMMARY

The human intestinal microbiota constitutes a complex ecosystem now well recognized for its impact on human health. It does contribute to prevention of colonization by pathogens and maturation of the immune system. Its possible implication in diseases of modern societies, currently increasing in prevalence, has been reported. These include allergies, inflammatory bowel diseases and possible obesity and cancer.

The analysis of the molecular composition of the human intestinal microbiota indicates marked inter-individual variations, which may seem paradoxical considering the high degree of conservation of major functions of the intestinal microbiota such as anaerobic digestion of alimentary fibres. We have characterized a phylogenetic core within the human intestinal microbiota, in terms of composition; i.e. a set of conserved species that could be responsible for major conserved functionalities. Based on culture independent molecular assessments, the current knowledge allows to define criteria qualifying the normal state of the human intestinal microbiota that we call "eubiosis". This further allows to identify specific distortion from eubiosis, i.e. dysbiosis in immune, metabolic or degenerative diseases. Noticeably, Crohn's disease, an inflammatory bowel disease of yet unknown aetiology, is associated with an intestinal dysbiosis with a lower representation of the Clostridium leptum group among the Firmicutes phylum. We further showed that the bacterial species Faecalibacterium prausnitzii is exerting anti-inflammatory properties in vitro and in animal models that could explain its ability, in vivo in patients, when it is detectable in the mucosa associated microbiota, to protect patients from post-operative recurrence of endoscopic signs of inflammation 6 months after surgical resection of the ileo-caecal region of the gut.

In confirming the major role of the microbiota in bowel related disorders, especially associated with a disruption of homeostasis, we are currently applying high throughput functional metagenomic screens in order to identify signal molecules and mechanisms of bacteria-host crosstalk. Together with the high resolution description of the human intestinal metagenome as well as explorations at the level of metaproteome and metabolome, these observations will further our understanding of the functional roles bacteria play in maintenance of health and well being in humans. It will open new perspectives for the monitoring and the design of strategies to modulate the microbiota for health.

INTRODUCTION

Microorganisms that colonize man from the very moment of birth on and establish with their host a mutualistic relationship that lasts for the whole life do play major roles that are well recognized today. Yet microorganisms that surround us are still most often perceived as dangerous potential invaders against which we should be ready to fight. This oddly negative perception contrasts with their acknowledged utility for environment, agriculture, industry and mankind.

The human organisms and before man, that of animals have long coevolved over millions of years with microorganisms, leading to the establishment of a mutually beneficial adaptation. Although fine mechanisms or temporal dynamics are not yet known, the host develops a tolerance towards non-pathogenic bacteria that colonize mucosal surfaces and skin and in return, the latter control the development and maintenance of numerous essential functions of their host. The intestinal microbiota contributes to bioconversion of food-born compounds unabsorbed in the upper parts of the digestive tract. Colonization by commensal microorganisms is key to immune development (Duarte et al., 2004; Neumann et al., 1998; Souza et al., 2004; Oliveira et al., 2005). The microbiota exerts in addition a direct role of colonization prevention keeping pathogens at levels of population preventing them from expressing virulence (Freter, 1983; Wells et al., 1988; van der Waaij et al., 1971). More recently, a potential role of the intestinal microbiota in regulation of fat storage has been described in mice (Backhed et al., 2005) and suggested to operate in man (Ley et al., 2006; Duncan et al., 2007).

Our vision of the human intestinal microbiota has recently been thoroughly revisited using molecular tools with a marker gene approach based on the use of ribosomal RNA as phylogenetic marker. For an overview of the main stages of the development of the complex microbiota associated to man the reader is invited to consult the appropriate chapter in this monograph. We herein will give an updated view of the intestinal microbiota composition and stability. We will thereafter outline its beneficial roles, and illustrate the association of microbiota dysbiosis and disease. We will finally give perspectives of application of metagenomic exploration of the functionalities of the dominant human intestinal microbiota.

COMPOSITION OF THE HUMAN INTESTINAL MICROBIOTA

Microbial population densities in the human digestive tract reach their maximum values in the colon with 10¹¹ bacteria per gram content. It is accepted today that this complex bacterial consortium known as the microbiota is made of hundreds of species in each individual. Yet a thorough description of all intestinal bacteria does not exist for two main reasons:

- 1. Traditional culture-based charac-
- terization (*Finegold* et al., 1974; *Holdeman* et al., 1976; *Moore* and *Holdeman*, 1974) does not allow to take into account more that 30% or so of the microorganisms that can be seen and enumerated by microscopic observation.
- 2. Species diversity of commensal intestinal bacteria at the level of the planet would be immense. In that respect, the use of molecular tools

indicated that the major part of dominant bacterial species observed in the faecal microbiota of an individual (approximately 80%) are specific of this individual (*Suau* et al., 1999; *Mangin* et al., 2004; *Eckburg* et al., 2005).

If species diversity of the dominant intestinal microbiota gives a faecal print essentially specific of the individual, composition at the level of taxa (genera and/or large phylogenetic groups) highlights consistent components, found in all individuals. Some of these taxa have been known for long and are represented by numerous collection strains; others have been evidenced only recently and via molecular approaches and still have no cultured representative. Culturable genera of the dominant faecal microbiota of adults are Bacteroides, Eubacterium, Ruminococcus, Clostridium and Bifidobacterium (Moore et al., 1974; Finegold et al., 1983). Accounting for non-culturable microorganisms allowed to refine this vision and to put it in a phylogenetic framework. The phylum Firmicutes is always highly represented. It comprises the Eubacterium rectale -Clostridium coccoides, often the most represented (14 to 31% of total bacteria depending on the studies) (Franks et al., 1998; *Jansen* et al., 1999; *Sghir* et al., 2000; Rigottier-Gois et al., 2003; Seksik et al., 2003). It is composed of species belonging to the genera Eubacterium, Clostridium, Ruminococcus, and Butyrivibrio. The phylum Firmicutes also comprises the Clostridium leptum group with the species Faecalibacterium prausnitzii, Ruminococcus albus and R. flavefaciens; a group that is also very often dominant (16 to 22%) on average) (Sghir et al., 2000; Lay et al., 2005). Bacteroidetes are represented by genera related to Bacteroides. They are always present and share

the dominance with the above groups (9 to 42% of total bacteria on average). The phylum Actinobacteria is less consistently detected in dominance, but represents a few percent of total bacteria. It comprises bifidobacteria (0.7 to 10%) and bacteria of the *Collinsella*-Atopobium group (0.3 to 3.7% on average) (Harmsen et al., 2000; Rigottier-Gois et al, 2003). Enterobacteria are more seldom observed in the top two logs of population in the faecal microbiota (0.4 to 1%), similarly to lactobacilli and streptococci (2%) (*Lay* et al., 2005). Also occasionally found are species related to Clostridium ramosum, Eubacterium cylindroids, Phascolarctobacterium, Verrucomicrobium or Sporomusa-Selenomonas-Veillonella.

Highly conserved composition traits at the level of phylogenetic groups and phyla on the one hand together with subject specificity at the level of species on the other hand suggest that there exists, on functional grounds, some degree of redundancy between species and the different levels of resolution bring complementary pieces of information.

Finally, bacterial species observed are strictly associated to the intestinal ecosystem. This derives from a long co-evolution with the host (*Ley* et al., 2006) that recent studies of cross-association of microbiota from/with different hosts do confirm (*Rawls* et al., 2006).

At present, phylogenetic reassessment of the human intestinal microbiota has been essentially restricted to the dominant fraction and our knowledge of subdominant bacteria (i.e. below 10⁸ per gram stool) may be incomplete and remains restricted to culturable isolates. The ability to isolate and grow microorganisms *in vitro* remains a key step in knowledge building, especially con-

sidering that phylogeny does not inform on the *in situ* activity of microbes. On a more focussed standpoint, the assessment of the contribution of archaea

or phages, that may be highly significant in terms of populations, has remained anecdotal up to now.

HOMEOSTASIS OF THE INTESTINAL MICROBIOTA

By definition, a microorganism that colonizes a given niche will persist and multiply without requiring re-inoculation. Dynamics and homeostasis of the intestinal microbiota may be considered in time (for any given individual) and space (i.e. between individuals or intestinal compartments). The global composition of the dominant intestinal microbial community appears conserved between individuals and with time. The same major phyla are present with proportions that vary between individuals but most likely remaining within the same log-unit equivalent in terms of population. Dominant species diversity appears remarkably stable with time for a given individual from day-to-day and even across years (Zoetendal et al., 98; VanHoutte et al., 2004; Seksik et al., 2003) while a large fraction of the dominant species appear specific of the subject. At the level of strains stability is more or less evident depending on the subject (McCartney et al., 1996; Kimura et al., 1997). Hence the stability observed at the level of groups and species would hide an important rate of renewal at the level of strains. Genomic plasticity may in fact come into play in that respect. It has also been shown that species diversity for subdominant groups (ex. Lactobacilus) is far less stable with time than that of dominant ones (VanHoutte et al., 2004) and that stability of communities is greater in the colon than in the ileum. It must also be considered that lactic acid bacteria brought by the ingestion of fermented food may occasionally have a survival rate during

transit that leads to their transient passage in dominance in the small intestine and colon.

For a given individual, modifications of the intestinal microbiota may derive either from colonization by exogenous microorganisms or by modulation of population levels of commensal bacteria. In most cases it will essentially be the consequence of relays in dominance, in response to factors modulating ecological niches. Numerous factors may affect stability of microbial communities - among others transit time, pH, quality and quantity of exogenous substrates and endogenous mucins. Although microbial communities appear ready to deal with changes in ecological settings, it seems difficult to induce durable alterations of established dominant populations, at least in terms of composition. Numerous observations hence illustrate the ability of the dominant intestinal microbiota to resist modification. The administration of an allochtonous strain such as a probiotic or an exogenous non-absorbable substrate such as a prebiotic often lead to transient modifications of microbial equilibrium. Even a major stress such as an antibiotic administration can be followed by a return of the community to its initial dominant species profile within a month or so (de la Cochetiere et al., 2005; *Dethlefsen* et al., 2008). This ability to recover its original make-up following a stress, known as resilience, suggests a fine-tuned adaptation of the microbiota to the gut and even to the host that harbours it. This can be linked to the observation that monozygotic twins have faecal microbial communities that have significantly more closely related patterns than these of unrelated individuals, suggesting that genotype may play a role in the development and structuration of the intestinal bacterial populations.

Analysing the spatial distribution of intestinal microbes as a function of digestive sites is difficult to study; it requires collection of samples within and along the intestine, hence via invasive methods. The preservation of topological relations between bacteria and epithelium is also a challenge. This explains some remaining controversy on this topic.

The luminal microbiota (within the intestinal cavity) has been explored in several ways. The proximal colon luminal microbiota differs from the faecal microbiota of which the composition only represents the distal parts of the colon (Marteau et al., 2001). Between the proximal and the distal colon, microbial populations increase overall by a factor of 100 and the increase is essentially due to an increase in strictly anaerobic bacteria. The layer of mucus that covers the intestinal wall constitutes a specific ecological niche. Several studies have shown that the microbial community that colonizes this niche is stable with time and remarkably comparable from the ileum to the rectum for a given individual (Lepage et al., 2005; Wang et al., 2005). Conversely, species that dominate in the mucus layer differ from dominant luminal species found in faeces (*Eckburg* et al., 2005; *Lepage* et al., 2005).

The ability of commensal gut bacteria to adhere *in situ* to intestinal epithelial cells has so far not been documented in an unequivocal manner. Indirect evidence does exist, derived from the presence of genes encoding

adhesions in the genome of strains of E. coli able to durably colonize their host. Adhesins could nonetheless contribute to the recognition of mucosal sites and structures or sloughed cells. In ecological terms, a given strain must divide at least as quickly as its offspring's are eliminated in order to maintain itself at a stable level of population in the ecosystem (*Lee* et al., 2004). Hence adhesion to the epithelium does not appear as an absolute necessity, but recognition of sites within the mucus or in the contents would provide a selective advantage for slow growing strains (Freter et al., 1983). If adhesion to the epithelium does not seem to be a relevant criterium for commensal bacteria, this property has been associated with intestinal bacteria in patients with inflammatory bowel diseases, and of course many intestinal pathogens.

It remains clear that not all mechanisms involved in maintenance of homeostasis of the human intestinal microbiota are understood to date; noticeably determinants of resistance to change and resilience. It hence is still fully relevant to question the right level of phylogenetic depth and time period for which stability of the ecosystem should be defined. As far as resilience is concerned, it is also reasonable to speculate that a certain level of stress will disturb the equilibrium of the gut ecosystem such that it will be irreversibly perturbed. The threshold above which the human intestinal microbiota loses its ability to return to its original balance is still unknown today.

Finally, parameters of homeostasis will apply to functionalities as much as composition; yet the relevance of these parameters on a functional standpoint is totally unexplored. Functional resistance and resilience of the intestinal microbiota have yet to be determined, as well as the link between phylogeny

and functions. There are only speculations at present on a potential link be-

tween "quantity of diversity" and functional resistance and resilience.

EUBIOSIS AND DYSBIOSIS OF THE HUMAN INTESTINAL MICROBIOTA

Based on culture independent molecular assessments, the current knowledge allows to define criteria qualifying the normal state of the human intestinal microbiota that we call "eubiosis". Obviously, emphasis has been put for technical reasons on phylogenetic evaluation of composition, diversity, core species and the dynamics of these over time and space and it is obvious that defining eubiosis will benefit from the addition of functional parameters. This context further allows to identify specific distortions from eubiosis, i.e. dysbiosis, which can be specifically investigated in immune, metabolic or degenerative diseases. We have recently validated the concept in the case of Crohn's disease. Crohn is an inflammatory bowel disease of yet unknown aetiology, that has a prevalence of one per 2000 in European countries. We have demonstrated that Crohn's disease is associated with an intestinal dysbiosis with a lower representation of the *Clostridium leptum* group among the Firmicutes phylum (Seksik et al., 2003; Sokol et al., 2006, 2008, 2009). We further showed that the bacterial species Faecalibacterium prausnitzii, when detectable in the mucosa associated microbiota of the ileum of patients, is protective against post-operative recurrence of endoscopic signs of inflammation 6 months after surgical resection of the ileo-caecal region of the intestine (Sokol et al., 2008). We finally demonstrated that Faecalibacterium prausnitzii could exert anti-inflammatory properties in vitro and in animal models with chemically induced inflammation.

The exploration of dysbiosis may be viewed as a primary step providing key information for the design of strategies aiming at restoring or maintaining homeostasis and eubiosis. Although so far restricted to microbiota composition and/or diversity, dysbiosis has been proposed and in a few cases well documented in irritable bowel syndrome (Kassinen et al., 2007), ulcerative colitis (Sokol et al., 2008; Martinez et al., 2008), obesity (Ley et al., 2007; Kalliomäki et al., 2008), Type-1 diabetes (*Dessein* et al., 2009; Wen et al., 2008), type-2 diabetes (Cani and Delzenne, 2009), celiac disease (Nadal et al., 2007; Collado et al. 2009), allergy (Kirjavainen et al., 2002; Björkstén, 2009), and in cases of infections with Clostridium difficile (Hickson et al., 2007) or HIV (Gori et al., 2008). These observations, just as that of dysbiosis in Crohn's disease above, are not indicative of a causal relationship between microbiota imbalance and onset of the disease. Indeed it is quite reasonable to argue that, once such diseases are declared, owing to the disruption they cause in the immune system and in physico-chemical properties of the intestinal milieu, dysbiosis could in fact be a consequence rather than a cause. In the case of Faecalibacterium prausnitzii in Crohn's disease, we nevertheless have a situation in which a deprivation in populations of a normal commensal bacterium, belonging to the most dominant core species of the healthy gut microbiota and potentially anti-inflammatory in vivo, will be associated with a reduced ability of the ecosystem to promote a return to immune

homeostasis. It can even be anticipated that a vicious circle is into place combining the detrimental effects of higher bacterial densities close to the mucosa (Swidsinski et al., 2005), increased populations of Gram-negative, pro-inflammatory, endotoxin producing bacteria usually subdominant in healthy subjects (Baumgart et al., 2007; Darfeuille-Michaud et al., 2004), reduced proportions of anti-inflammatory commensals (Sokol et al., 2008) and even increased occurrence of protein biomarkers, potentially promoting auto-

immune reactivity (Juste and Doré, personal communication).

The current strengthening of the concept of eubiosis/dysbiosis confirms the major role of the microbiota in bowel related disorders, especially associated with a disruption of homeostasis. It stresses the need to apply the emerging tools of microbiomics to provide diagnostic models and also to identify signal molecules and describe bacteria-host crosstalk mechanisms at play.

HUMAN INTESTINAL MICROBIOMICS

Beyond the phylogenetic level extensively explored so far lie the combined genomes, transcriptomes, proteomes and even metabolomes of the members of the intestinal microbial community. These are known today as the metagemetatranscriptome, metaproteome, .. and all-together the human intestinal microbiome. Since the vast majority of dominant intestinal bacteria are not yet cultured to date, the genomic content of this microbial community and its derived components at the various levels of omic integration are essentially unknown. Metagenomics is emerging today as this most powerful approach to characterize the repertoire of genes of any complex microbial setting, independent of the culturability of its components. The development of very high throughput sequencing technologies is further contributing to this development. In practical terms, the microbial community may be extracted from its environment and its DNA purified and/or cloned in order to determine its sequence. The complete gene repertoire of culturable and non-culturable dominant microbes can hence be obtained. This further offers the possibility to design high

throughput profiling tools informative at the level of functional potentials of the complete community. In addition, cloning of genome fragments of intestinal bacteria in large insert metagenomic libraries allows to use functional screens in order to seek functions of non-culturable bacteria after heterologous expression in E. coli, giving access to yet totally unexplored biological resources. Beyond its innovative character, the potentials of metagenomics have already largely been documented (Riesenfeld et al., 2004; The new science of metagenomics: revealing the secrets of our microbial planet, National Research Council of the National Academies, The National Academies Press, Washington DC, USA, 2007).

The first developments of the metagenomics approach applied to the human intestinal microbiota have focussed on diversity of the microbial community (Manichanh et al., 2006, 2008) and the gene repertoire for a few subjects (Gill et al., 2006; Kurokawa et al., 2007). Functional metagenomics applications have remained confined to the soil ecosystem (Williamson et al., 2005) and animal guts (Beloqui et al.,

2006). We initiated its application to the human intestinal context (*Gloux* et al., 2007). Although still in its infancy, this approach brings major promises for an improved understanding of microbe-food, microbe-host and microbe-microbe interactions.

The microbiomic exploration of the human intestinal microbiota is hence ongoing thanks to several programs such as the European Commission-funded program MetaHIT (http://locus.jouy.inra.fr/metahit/), and the French Agency for Research-funded program GMGE Micro-Obes (http://www.inra.fr/micro_obes), as well as the NIH Roadmap programs (http://nihroadmap.nih.gov/hmp/) and

many others worldwide. At the international level these programs are structured within the International Human Microbiome Consortium (IHMC) co-chaired by the NIH and the European Commission. These programs will deliver a huge mass of information that will in turn allow to identify conserved and variable genomic and functional traits of the ecosystem, to describe those specific to the gut environment and bearing the best diagnostic and/or prognostic potential, to reconstruct the metabolic food-chain of the microbial community, and to start describe ecotypes and model their relationships in a systems' ecology endeavour.

CONCLUSION

Application of molecular ecology tools to the intestinal microbiota has allowed very significant improvements in our understanding of this ecosystem in terms of composition and dynamics of species diversity. The single gene approach based on ribosomal RNA as a universal phylogenetic marker had nevertheless left aside the functions microorganisms exert in their environment.

It has become possible to sequence combined genomes of complex microbial communities giving access to their potential activities. The following steps towards environmental transcriptomics, expressed proteins, activities and metabolites are being taken. The global functional exploration of the human intestinal microbiota is hence underway, with perspectives as large and fascinating as these of the former decade.

LITERATURE

Backhed, F., Ley, R.E., Sonnenburg, J.L., Peterson, D.A., and Gordon, J.I.: Host-bacterial mutualism in the human intestine. Science 307, 1915-1920 (2005).

Baumgart, M., Dogan, B., Rishniw, M.,
Weitzman, G., Bosworth, B., Yantiss, R.,
Orsi, R.H., Wiedmann, M., McDonough,
P., Kim, S.G., Berg, D., Schukken, Y.,
Scherl, E., and Simpson, K.W.: Culture independent analysis of ileal mucosa reveals
a selective increase in invasive *Escherichia*

coli of novel phylogeny relative to depletion of Clostridiales in Crohn's disease involving the ileum. ISME J. 1, 403-418 (2007).

Beloqui, A., Pita, M., Polaina, J., Martínez-Arias, A., Golyshina, O.V., Zumárraga, M., Yakimov, M.M., García-Arellano, H., Alcalde, M., Fernández, V.M., Elborough, K., Andreu, J.M., Ballesteros, A., Plou, F.J., Timmis, K.N., Ferrer, M., and Golyshin, P.N.: Novel polyphenol oxidase

- mined from a metagenome expression library of bovine rumen: Biochemical properties, structural analysis, and phylogenetic relationships. Biol. Chem. 281, 22933-22942 (2006).
- Björkstén, B.: Disease outcomes as a consequence of environmental influences on the development of the immune system. Curr. Opin. Allergy Clin. Immunol. 9, 185-189 (2009).
- Cani, P.D. and Delzenne, N.M.: The role of the gut microbiota in energy metabolism and metabolic disease. Curr. Pharm. Des. 15, 1546-1558 (2009).
- Collado, M.C., Donat, E., Ribes-Koninckx, C., Calabuig, M., and Sanz, Y.: Specific duodenal and faecal bacterial groups associated with paediatric coeliac disease. J. Clin. Pathol. 62, 264-269 (2009).
- Darfeuille-Michaud, A., Boudeau, J., Bulois, P., Neut, C., Glasser, A.L., Barnich, N., Bringer, M.A., Swidsinski, A., Beaugerie, L., and Colombel, J.F.: High prevalence of adherent-invasive *Escherichia coli* associated with ileal mucosa in Crohn's disease. Gastroenterology 127, 412-421 (2004).
- De la Cochetière, M.-F., Durand, T., Lepage, P., Bourreille, A., Galmiche, J.-P., and Doré, J.: Resilience of the dominant human fecal microbiota upon shortcourse antibiotic challenge. J. Clin. Microbiol. 43, 5588-5592 (2005).
- Dessein, R., Peyrin-Biroulet, L., and Chamaillard, M.: Intestinal microbiota gives a nod to the hygiene hypothesis in type 1 diabetes. Gastroenterology 137, 381-383 (2009).
- Dethlefsen, L., Huse, S., Sogin, M.L., and Relman, D.A.: The pervasive effects of an antibiotic on the human gut microbiota, as revealed by deep 16S rRNA sequencing. PLoS Biol. 6, e280 (2008).
- Duarte, R., Silva, A.M., Vieira, L.Q., Alfonso, L.C., and Nicoli, J.R.: Influence of normal microbiota on some aspects of the immune response during experimental infection with *Trypanosoma cruzi* in mice. J. Med. Microbiol. 53, 741-748 (2004).
- Duncan, S.H., Belenguer, A., Holtrop, G., Johnstone, A.M., Flint, H.J., and Lobley,

- G.E.: Reduced dietary intake of carbohydrates by obese subjects results in decreased concentrations of butyrate and butyrate-producing bacteria in feces. Appl. Environ. Microbiol. 73, 1073-1078 (2007).
- Eckburg, P.B., Bik, E.M., Bernstein, C.N., Purdom, E., Dethlefsen, L., Sargent, M., Gill, S.R., Nelson, K.E., and Relman, D.A.: Diversity of the human intestinal microbial flora. Science 308, 1635-1638 (2005).
- Finegold, S.M., Attebery, H.R., and Sutter, V.L.: Effect of diet on human fecal flora: Comparison of Japanese and American diets. Am. J. Clin. Nut. 27, 1456-1469 (1974).
- Franks, A.H., Harmsen, H.J., Raangs, G.C., Jansen, G.J., Schut, F., and Welling, G.W.: Variations of bacterial populations in human feces measured by fluorescent *in situ* hybridization with group-specific 16S rRNA-targeted oligonucleotide probes. Appl. Environ. Microbiol. 64, 3336-3345 (1998).
- Freter, R.: Mechanisms that control the microflora in the large intestine. In: Human intestinal microflora in health and disease (Hentges, D.J., Ed.). Academic Press, New York/London, 33-54 (1983).
- Freter, R., Brickner, H., Fekete, J., Vickerman, M.M., and Carey, K.E.: Survival and implantation of *Escherichia coli* in the intestinal tract. Infect. Immun. 39, 686-703 (1983).
- Gill, S.R., Pop, M., Deboy, R.T., Eckburg, P.B., Turnbaugh, P.J., Samuel, B.S., Gordon, J.I., Relman, D.A., Fraser-Liggett, C.M., and Nelson, K.E.: Metagenomic analysis of the human distal gut microbiome. Science 312, 1355-1359 (2006).
- Gloux, K., Leclerc, M., Iliozer, H., L'Haridon, R., Manichanh, C., Corthier, G., Nalin, R., Blottière, H.M., and Doré, J.: Development of high-throughput phenotyping of metagenomic clones from the human gut microbiome for modulation of eukaryotic cell growth. Appl. Environ. Microbiol. 73, 3734-3737 (2007).
- Gori, A., Tincati, C., Rizzardini, G., Torti, C.,

- Quirino, T., Haarman, M., Ben Amor, K., van Schaik, J., Vriesema, A., Knol, J., Marchetti, G., Welling, G., and Clerici, M.: Early impairment of gut function and gut flora supporting a role for alteration of gastrointestinal mucosa in human immunodeficiency virus pathogenesis. J. Clin. Microbiol. 46, 757-758 (2008).
- Harmsen, H.J.M., Wildeboer-Veloo, A.C.M., Grijpstra, J., Knol, J., Degener, J.E., and Welling, G.W.: Development of 16S rRNA-based probes for the Coriobacterium group and the Atopobium cluster and their application for enumeration of Coriobacteriaceae in human feces from volunteers of different age groups. Appl. Environ. Microbiol. 66, 4523-4527 (2000).
- Hickson, M., D'Souza, A.L., Muthu, N., Rogers, T.R., Want, S., Rajkumar, C., and Bulpitt, C.J.: Use of probiotic Lactobacillus preparation to prevent diarrhoea associated with antibiotics: Randomised double blind placebo controlled trial. BMJ 335, 380 (2007).
- Holdeman, L.V., Good, I.J., and Moore, W.E.C.: Human fecal flora: Variation in bacterial composition within individuals and a possible effect of emotional stress. Appl. Environ. Microbiol. 31, 359-375 (1976).
- Jansen, G.J., Wildeboer-Veloo, A.C., Tonk, R.H., Franks, A.H., and Welling, G.W.: Development and validation of an automated, microscopy-based method for enumeration of groups of intestinal bacteria. J. Microbiol. Methods 37, 215-221 (1999).
- Kalliomäki, M., Isolauri, E., and Salminen, S.: Probiotics and prevention of atopic disease: 4-year follow-up of randomized placebo-controlled trial. Lancet 361, 1869-1871 (2003).
- Kalliomäki, M., Collado, M.C., Salminen, S., and Isolauri, E.: Early differences in fecal microbiota composition in children may predict overweight. Am. J. Clin. Nutr. 87, 534-538 (2008).
- Kassinen, A., Krogius-Kurikka, L., Mäkivuokko, H., Rinttilä, T., Paulin, L., Corander, J., Malinen, E., Apajalahti, J., and Palva,

- A.: The fecal microbiota of irritable bowel syndrome patients differs significantly from that of healthy subjects. Gastroenterology 133, 24-33 (2007).
- Kimura, K. McCartney, A.L., McConnell, M.A., and Tannock, G.W.: Analysis of fecal populations of bifidobacteria and lactobacilli and investigation of the immunological responses of their human hosts to the predominant strains. Appl. Environ. Microbiol. 63, 3394-3398 (1997).
- Kirjavainen, P.V., Arvola, T., Salminen, S.J., and Isolauri, E.: Aberrant composition of gut microbiota of allergic infants: A target of bifidobacterial therapy at weaning? Gut 51, 51-55 (2002).
- Kurokawa, K., Itoh, T., Kuwahara, T., Oshima, K., Toh, H., Toyoda, A., Takami, H., Morita, H., Sharma, V.K., Srivastava, T.P., Taylor, T.D., Noguchi, H., Mori, H., Ogura, Y., Ehrlich, S.D., Itoh, K., Takagi, T., Sakaki, Y., Hayashi, T., and Hattori, M.: Comparative metagenomics revealed commonly enriched gene sets in human gut microbiomes. DNA Res. 14, 169-181 (2007).
- Lay, C., Sutren, M., Rochet, V., Saunier, K., Doré, J., and Rigottier-Gois, L.: Design and validation of 16S rRNA probes to enumerate members of the *Clostridium leptum* subgroup in human faecal microbiota. Environ. Microbiol 7, 933–946 (2005).
- Lee, Y.K., Ho, P.S., Low, C.S., Arvilommi, H., and Salminen, S.: Permanent colonization by *Lactobacillus casei* is hindered by low rate of cell division in mouse gut. Appl. Environ. Microbiol. 70, 670-674 (2004).
- Lepage, P., Seksik, P., Sutren, M., de la Cochetiere, M.F., Jian, R., Marteau, P., and Doré, J.: Biodiversity of the mucosa-associated microbiota is stable along the distal digestive tract in healthy individuals and patients with IBD. Inflamm. Bowel. Dis. 11, 473-480 (2005).
- Ley, R.E., Backhed, F., Turnbaugh, P., Lozupone, C.A., Knight, R.D., and Gordon, J.I.: Obesity alters gut microbial ecology. Proc. Natl. Acad. Sci. USA 102, 11070-11075 (2005).

- Ley, R.E., Peterson, D.A., and Gordon, J.I.: Ecological and evolutionary forces shaping microbial diversity in the human intestine. Cell 124, 837-848 (2006).
- Mangin, I., Bonnet, R., Seksik, P., Rigottier-Gois, L., Sutren, M., Bouhnik, Y., Neut, C., Collins, M.D., Colombel, J.-F., Marteau, P., and Doré, J.: Molecular inventory of faecal microflora in patients with Crohn's disease. FEMS Microbiol. Ecol. 50, 25-36 (2004).
- Manichanh, C., Rigottier-Gois, L., Bonnaud, E., Gloux, K., Pelletier, E., Frangeul, L., Nalin, R., Jarrin, C., Chardon, P., Marteau, P., Roca, J., and Doré J.: Reduced diversity of faecal microbiota in Crohn's disease revealed by a metagenomic approach. Gut 55, 205-211 (2006).
- Manichanh, C., Chapple, C.E., Frangeul, L., Gloux, K., Guigo, R., and Dore, J.: A comparison of random sequence reads versus 16S rDNA sequences for estimating the biodiversity of a metagenomic library. Nucleic Acids Res. 36, 5180-5188 (2008).
- Marteau, P., Pochart, P., Doré, J., Maillet, C., Bernalier, A., and Corthier, G.: Comparative study of the human cecal and fecal flora. Appl. Environ. Microbiol. 67, 4939-4942 (2001).
- Martinez, C., Antolin, M., Santos, J., Torrejon, A., Casellas, F., Borruel, N., Guarner, F., and Malagelada, J.R.: Unstable composition of the fecal microbiota in ulcerative colitis during clinical remission. Am. J. Gastroenterol. 103, 643-648 (2008).
- McCartney, A.L., Wenzhi, W., and Tannock, G.W.: Molecular analysis of the composition of the bifidobacterial and lactobacillus microflora of humans. Appl. Environ. Microbiol. 62, 4608-4613 (1996).
- Moore, W.E.C. and Holdeman, L.V.: Human fecal flora: The normal flora of 20 Japanese-Hawaiians. Appl. Microbiol. 27, 961-979 (1974).
- Nadal, I., Donat, E., Ribes-Koninckx, C., Calabuig, M., and Sanz, Y.: Imbalance in the composition of the duodenal microbiota of children with coeliac disease. J. Med. Microbiol. 56, 1669-1674 (2007). Erratum

- in: J. Med. Microbiol. 57, 401 (2008).
- Neumann, E., Oliveira, M.A., Cabral, C.M., Moura, L.N., Nicoli, J.R., Vieira, E.C., Cara, D.C., Podoprigora, G.I., and Vieira, L.Q.: Monoassociation with *Lactobacillus* acidophilus UFV-H2b20 stimulates the immune defense mechanisms of germfree mice. Braz. J. Med. Biol. Res. 31, 1565-1573 (1998).
- Oliveira, M.R., Tafuri, W.L., Afonso, L.C., Oliveira, M.A., Nicoli, J.R., Vieira, E.C., Scott, P., Melo, M.N., and Vieira, L.Q.: Germfree mice produce high levels of interferon-gamma in response to infection with *Leishmania major* but fail to heal lesions. Parasitology 131, 477-488 (2005).
- Rawls, J.F., Mahowald, M.A., Ley, R.E., and Gordon, J.I.: Reciprocal gut microbiota transplants from zebrafish and mice to germ-free recipients reveal host habitat selection. Cell 127, 423-433 (2006).
- Riesenfeld, C.S., Schloss, P.D., and Handelsman, J.: Metagenomics: Genomic analysis of microbial communities. Annu. Rev. Genet. 38, 525-552 (2004).
- Rigottier-Gois, L., Le Bourhis, A.G., Gramet, G., Rochet, V., and Doré, J.: Fluorescent hybridisation combined with flow cytometry and hybridisation of total RNA to analyse the composition of microbial communities in human faeces using 16S rRNA probes. FEMS Microbiol. Ecol. 43, 237-245 (2003).
- Seksik, P., Rigottier-Gois, L., Gramet, G., Sutren, M., Pochart, P., Marteau, P., Jian, R., and Doré, J.: Alterations of the dominant faecal bacterial groups in patients with Crohn's disease of the colon. Gut 52, 237-242 (2003).
- Sghir, A., Gramet, G., Suau, A., Rochet, V., Pochart, P., and Doré, J.: Quantification of bacterial groups within human fecal flora by oligonucleotide probe hybridization. Appl. Environ. Microbiol. 66, 2263-2266 (2000).
- Sokol, H., Seksik, P., Rigottier-Gois, L., Lay, C., Lepage, P., Podglajen, I., Marteau, P., and Doré, J.: Specificities of the fecal microbiota in inflammatory bowel disease.

- Inflamm. Bowel Dis. 12, 106-111 (2006).
- Sokol, H., Pigneur, B., Watterlot, L., Lakhdari, O., Bermúdez-Humarán, L.G., Gratadoux, J.J., Blugeon, S., Bridonneau, C., Furet, J.P., Corthier, G., Grangette, C., Vasquez, N., Pochart, P., Trugnan, G., Thomas, G., Blottière, H.M., Doré, J., Marteau, P., Seksik, P., and Langella, P.: Faecalibacterium prausnitzii is an anti-inflammatory commensal bacterium identified by gut microbiota analysis of Crohn disease patients. Proc. Natl. Acad. Sci. USA 105, 16731-16736 (2008).
- Sokol, H., Seksik, P., Furet, J.P., Firmesse, O., Nion-Larmurier, I., Beaugerie, L., Cosnes, J., Corthier, G., Marteau, P., and Doré, J.: Low counts of *Faecalibacterium praus-nitzii* in colitis microbiota. Inflamm. Bowel Dis. 15, 1183-1189 (2009).
- Souza, D.G., Vieira, A.T., Soares, A.C., Pinho, V., Nicoli, J.R., Vieira, L.Q., and Teixeira, M.M.: The essential role of the intestinal microbiota in facilitating acute inflammatory responses. J. Immunol. 173, 4137-4146 (2004).
- Suau, A., Bonnet, R., Sutren, M., Godon, J.J., Gibson, G.R., Collins, M.D., and Doré, J.: Direct analysis of genes encoding 16S rRNA from complex communities reveals many novel molecular species within the human gut. Appl. Environ. Microbiol. 65, 4799-4807 (1999).
- Swidsinski, A., Weber, J., Loening-Baucke, V., Hale, L.P., and Lochs, H.: Spatial organization and composition of the mucosal flora in patients with inflammatory bowel disease. J. Clin. Microbiol. 43, 3380-3389 (2005).
- van der Waaij, D., de Vries, B., and van der Wees, L.: Colonization resistance of the

- digestive tract in conventional and antibiotic-treated mice. J. Hyg. 69, 405-411 (1971).
- Vanhoutte, T., Huys, G., De Brandt, E., and Swings, J.: Temporal stability analysis of the microbiota in human feces by denaturing gradient gel electrophoresis using universal and group-specific 16S rRNA gene primers. FEMS Microbiol. Ecol. 48, 437-446 (2004).
- Wang, M., Ahrné, S., Jeppsson, B., and Molin, G.: Comparison of bacterial diversity along the human intestinal tract by direct cloning and sequencing of 16S rRNA genes. FEMS Microbiol. Ecol. 54, 219-231 (2005).
- Wells, C.L., Maddaus, M.A., and Simmons, R.L.: Proposed mechanisms for the translocation of intestinal bacteria. Rev. Infect. Dis. 10, 958-979 (1988).
- Wen, L., Ley, R.E., Volchkov, P.Y., Stranges, P.B., Avanesyan, L., Stonebraker, A.C., Hu, C., Wong, F.S., Szot, G.L., Bluestone, J.A., Gordon, J.I., and Chervonsky, A.V.: Innate immunity and intestinal microbiota in the development of Type 1 diabetes. Nature 455, 1109-1113 (2008).
- Williamson, L.L., Borlee, B.R., Schloss, P.D., Guan, C., Allen, H.K., and Handelsman, J.: Intracellular screen to identify metagenomic clones that induce or inhibit a quorum-sensing biosensor. Appl. Environ. Microbiol. 71, 6335-6344 (2005).
- Zoetendal, E.G., Akkermans, A.D., and de Vos, W.M.: Temperature gradient gel electrophoresis analysis of 16S rRNA from human fecal samples reveals stable and host-specific communities of actives bacteria. Appl. Environ. Microbiol. 64, 3854-3859 (1998).